

Original article:

Bacteriological profile and susceptibility pattern of burn wound isolates in a tertiary care hospital

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Abstract

Burns patients are highly susceptible for opportunistic infections. A retrospective study was conducted to analyse the bacterial isolates from burn wound.

Keywords: Antibiotic susceptibility Burns, Infection

Introduction

Burn patients are highly susceptible for opportunistic infections^[1]. Infection is a major problem in the management of burn patients. Approximately 75% of mortality following burns are because of infection^[2,3,4]. Burn wound monitoring requires the study of changing bacterial flora and their antibiotic susceptibility pattern.^[5] This will help to assess the organisms that are predominant in a particular treatment centre and antimicrobial susceptibility testing will help to formulate antibiotic policy for better management of these patients. The present study is undertaken with the following aims and objectives:

1. To find out the bacterial profile for post burn infection in pus and blood.
2. To evaluate the antibiotic sensitivity of organisms cultured and isolated.

Material and method

This is a retrospective study of bacterial isolates from 200 wound swabs taken from 179 patients admitted to the burn unit of a tertiary care hospital in Mangalore. The specimens were transported in sterile,

leak-proof containers to the Microbiology department. All specimens were inoculated on 5% blood agar, MacConkey agar and Chocolate agar plates and incubated overnight at 37°C aerobically. Bacterial pathogens were identified by conventional biochemical methods according to standard microbiological techniques.

Antimicrobial susceptibility was performed on Mueller-Hinton agar by the standard disk diffusion method recommended by the The antibiotics tested for gram-positive cocci were: Ampicillin (10 mg), Cefoxitin (30 mg), Ceftriaxone (30 mg), Ciprofloxacin (5 mg), Azithromycin (15 mg), Vancomycin (30 mg), Linezolid (30 mg); for gram-negative bacilli: Ampicillin (10 mg), Amikacin (30 mg), Gentamicin (30 mg), Ciprofloxacin (5 mg), Piperacillin/Tazobactam (30/10 mg), Cefoperazone/Sulbactam (75/30 mg), Imipenem (10 mg) and for non-fermenters ceftazidime, (30 mg), piperacillin (100 mg), cefepime (30 mg), amikacin (30 mg), gentamicin (30 mg), ciprofloxacin (5 mg), cefoperazone / sulbactam (75/30 mg), Piperacillin/Tazobactam (75 /30 mg), Meropenem (10 mg) and

Imipenem (10 mg) were used. The antimicrobial susceptibilities were determined according to the Clinical and Laboratory Standards Institute (CLSI) guidelines. Extended spectrum beta lactamase (ESBL) production was tested by double disk approximation method. Methicillin resistant Staphylococcus aureus was screened using Cefoxitin (30 mg) disk.

Results

A total of 230 bacterial isolates were obtained from 179 patients' wound swab over a 3-year period. The most predominant bacterial isolate was *Acinetobacter* spp (34 %) , *Pseudomonas aeruginosa* (*P. aeruginosa*) (29%), *Klebsiella pneumonia* (22%) *Staphylococcus aureus* (*S. aureus*) (8%), *Escherichia coli* (*E. coli*) (1%) and *Enterococcus* spp (4%) as shown in Table 1.

Table 1/Fig 1

Distribution of microorganisms isolated from burns wound

Microorganism	n	%
Acinetobacter spp	78	34
Pseudomonas aeruginosa	68	29
Klebsiella pneumonia	51	22
Staphylococcus aureus	19	8
Enterococcus spp	9	4
Escherichia coli	4	1

Table 2/Fig 2

Susceptibilities of gram negative isolates to various antimicrobials

Antimicrobial	Acinetobacter spp		Pseudomonas spp		Klebsiella spp		Escherichia coli	
	N *	R (%) †,	N	R (%)	n	R (%)	n	R (%)
Ceftazidime	NT ‡	-	68	36 (52.9)	NT	-	NT	-
Piperacillin	NT	-	68	43 (63.2)	NT	-	NT	-
Ceftriaxone	78	74 (94.8)	68	53 (77.9)	51	36 (70.5)	4	0
Gentamicin	78	67 (85.8)	68	42 (61.7)	51	32 (62.7)	4	2 (50)
Amikacin	78	62 (79.4)	68	40 (58.8)	51	29 (56.8)	4	1 (25)
Ciprofloxacin	78	72 (92.3)	68	45 (66.1)	51	14 (27.4)	4	1 (25)

Imipenam	78	32 (41)	68	16 (23.5)	51	6 (11.7)	4	0
Meropenem	78	30 (38.4)	68	18 (26.4)	51	5 (9.8)	4	0
Cefoperazone +Salbactum	78	43 (55.1)	68	22(32.3)	51	6 (11.7)	4	1 (25)
Piperacillin + Tazobactum	78	37(47.4)	68	21 (30.8)	51	7 (13.7)	4	2 (50)
Tigecycline	78	2(2.5)	68	0	51	0	4	0
Colistin	78	0	68	0	NT	-	NT	-

N: number of isolates which were tested, R: resistant, NT: not tested

Table 3 /Fig 3

Susceptibilities of gram positive isolates to various antimicrobials

Antimicrobial	Staphylococcus aureus		Enterococcus spp	
	N	R (%)	N	R (%)
Ampicillin	19	16 (84.2)	9	2 (22.22)
Cefoxitin	19	5 (26.3)	NT	-
Ceftriaxone	19	5 (26.3)	NT	-
Gentamicin	19	09 (47.3)	NT	-
Amikacin	19	09 (47.3)	9	3 (33.33)
Ofloxacin	19	10 (52.6)	9	2 (22.22)
Vancomycin	19	0	9	0
Linezolid	19	0	9	0
Azithromycin	19	07 (36.8)	9	5 (55.55)

Acinetobacter spp was the most predominant organism isolated from the burns wound. Followed by Pseudomonas aeruginosa and Klebsiella pneumoniae. Among all the Acinetobacter spp and Pseudomonas spp 38% and 23% respectively were multidrug resistant. Tigecycline and Colistin were the only antimicrobial active against these isolates. Among all the Enterobacteriaceae 42% of Klebsiella and 26% of E.coli were extended spectrum beta lactamase (ESBL) producers. The most effective

antimicrobial against Klebsiella and E.coli was Meropenem.

Among the S. aureus isolated from patients within the burn center, the incidence of methicillin-resistant S. aureus (MRSA) was 26% and the most active antimicrobial agents were found to be Vancomycin and Linezolid against S. aureus isolates respectively. None of the Enterococcus spp. was found to be resistant to Vancomycin.

Discussion

Colonisation of burn wounds with microorganisms is almost certain to occur in patients with major burns. The most obvious reason is the unparalleled excellence of the burn wound as a bacterial microbiological culture medium. Devitalised tissues serves as a pabulum for microbial growth and with decreased blood flow to burn wound as a contributing factor^[6].

The most commonly isolated organism in the present study was *Acinetobacter baumannii* followed by *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*. *Acinetobacter* spp was shown to be an important cause of nosocomial infection in burns unit^[7]. Several studies have reported it to a predominant organism in burns unit. Some of the reasons for this trend is its presence as a normal commensal and easy spread due to multidrug resistance in hospital settings^[8,9]. *Pseudomonas aeruginosa* was isolated from 29% of wound swabs. This is in contrast to other studies where *Pseudomonas aeruginosa* was the commonest isolate^[10]. *Klebsiella pneumoniae* was the third in line. However its frequency is higher as compared to other studies^[11,12]. The isolation rate of gram negative bacilli was higher as compared to the gram positive cocci. This is in contrast to other studies where the isolation rate of *Staphylococcus aureus* was much higher^[13,14]. Amongst the gram

positive cocci *Staphylococcus aureus* was predominant followed by *Enterococcus* spp.

Acinetobacter spp and *Pseudomonas aeruginosa* were multidrug resistant with only Tigecycline and colistin active against these organisms. Singh et al. also reported high prevalence of multidrug resistant gram negative bacilli in burns unit. In this study Carbapenems like Imipenem and Meropenem and combination drugs like Piperacillin/Tazobactam and Cefoperazone/Salbactam showed good efficacy. This is in agreement with other studies. One of the reasons stated by Mehta et al. was that these third generation drugs are used as reserve drugs^[15,16]. ESBL production was lesser as compared to other studies^[9]. Guggenheim *et al* have showed that Imipenem and Meropenem were the most active antimicrobial agents for ESBL producing strains^[17]. Our results were in agreement with this study. MRSA was found among 26 % of the *Staphylococcus aureus* isolates which was lesser in comparison to other studies^[18,19]. MRSA isolated in this study were sensitive to Vancomycin, Linezolid and Amikacin.

Conclusion:

In conclusion high antimicrobial resistance is a major concern in burns unit. Aggressive infection control measures, judicious use of antibiotics as per the prevailing antibiotic susceptibility patterns should be applied to limit the emergence and spread of multidrug-resistant pathogens.

References

1. Cochran A, Morris SE, Edelman LS, Saffle JR, 2002. Systemic Candida infection in burn patients. Surg Infect-Larch mt. Vol 3 (4). pp367-74.
2. Macedo JLS, Santos JB. Bacterial and fungal colonization of burn wounds. Mem Inst Oswaldo Cruz 2005 ;100:535-39.
3. Taneja N, Emmanue IR, Chari PS, Sharma M. A prospective study of hospital acquired infections in burn patients at a tertiary care referral centre in North India. Burns 2004;30:248-53.

4. Vindenal H & Bjerknes R .Microbial colonization of large wounds.Burns 1995;21:575-79.
5. Bairy I, Shivananda PG. Aerobic bacterial flora of burn wound infection. Indian J Surg. 1997;59:215–8.
6. Church D,Elsayed S,Reid O,Winston B &Lindsay R.Burn wound infections. Clin.Microbiol. Rev.2006; 19 :40 3-34.
7. Sengupta S,Kumar P,CirajAM,Sivananda PG.Acinetobacter baumannii – an emerging nosocomial pathogen in the burns unit .Manipal,India.Burns 2001;27:140-44.
8. Chim H,Tan BH,Song C.Five year review of infections in a burn intensive care unit : High incidence of Acinetobacter baumannii in a tropical climate .Burns .2007;33(8):1008-14.
9. Bayram Y,Parlak M,Aypak C,Bayram I.Three year review of bacteriological profile and antibiogram of burn wound isolates in Van Turkey .Int J Med Sci.2013 ;10(1):19-23.
10. Ulku A,Serpil E,Mufide AN,Fehmi C,Ayten K.The time related changes of antimicrobial resistance patterns and predominant bacterial profiles of burns wounds and body flora of burned patients .Burns .2004-;30: 660-64.
11. Vindenes HA,Ulvested E,Bjerkenes R.Concentration of cytokines in plasma of patients with large burns ; their relation to time after injury,burn size,inflammatory variables,infection &outcome.Eur J Surg. 1998; 16 4(9):647-56.
12. Revathi G, Puri J, Jain BK. Bacteriology of burns. *Burns*. 1998;24(4):347-9 .
13. Dhar S,Saraf R,Singh K ,Raina B.Microbiological profile of chronic burn wounds among patients admitted to Santucci SG, Gobara S, Santos CR, Fontana C, Levin AS. Infections in a burn intensive care unit : experience of seven years. J Hosp Infect 2003; 53, 6-13.
14. Singh NP,Goyal R,Manchanda V,Das S,Kaur Z,Talwar V.Changing trends in the bacteriology of burns in the burns unit ,Delhi ,India .Burns .2003;29:129-32.
15. Mehta M,Dutta P,Gupta V.Bacterial isolates from burn wound infections and their antibiograms : A eight year study .Indian J Plast Surg 2007;40:25-28.
16. Guggenheim M, Zbinden R, Handschin AE, Gohritz A, Altintas MA, Giovanoli P. Changes in bacterial isolates from burn wounds and their antibiograms: a 20-year study (1986-2005). *Burns*. 2009;35(4):553-60
17. Buzaid N,Elzouki AN,Taher I,Ghengesh KS.Methicillin resistant Staphylococcus aureus (MRSA) in a tertiary surgical and trauma hospital in Benghazi,Libya.J Infect Dev Ctries .2011 ; 5(10):723-26.
18. Zorgani A,Shawref O ,Tawil K ,El –Turki E,Ghengesh KS.Inducible clindamycin resistance among staphylococci isolated from burn patients .Libyan J.